Synthesis, Characterization, and Utilization of a Lignin-Based Adsorbent for Effective Removal of Azo Dye from Aqueous Solution

Xianzhi Meng,* Brent Scheidemantle, Mi Li, Yun-yan Wang, Xianhui Zhao, Miguel Toro-González, Priyanka Singh, Yunqiao Pu, Charles E. Wyman, Soydan Ozcan, Charles M. Cai, and Arthur J. Ragauskas*

ABSTRACT: How to effectively remove toxic dyes from the industrial wastewater using a green low-cost lignocellulose-based adsorbent, such as lignin, has become a topic of great interest but remains quite challenging. In this study, cosolvent-enhanced lignocellulosic fractionation (CELF) pretreatment and Mannich reaction were combined to generate an aminated CELF lignin which is subsequently applied for removal of methylene blue and direct blue (DB) 1 dye from aqueous solution. $^{31}$P NMR was used to track the degree of amination, and an orthogonal design was applied to determine the relationship between the extent of amination and reaction parameters. The physicochemical, morphological, and thermal properties of the aminated CELF lignin were characterized to confirm the successful grafting of diethylenetriamine onto the lignin. The aminated CELF lignin proved to be an effective azo dye-adsorbent, demonstrating considerably enhanced dye decolorization, especially toward DB 1 dye (>90%). It had a maximum adsorption capacity of DB 1 dye of 502.7 mg/g, and the kinetic study suggested the adsorption process conformed to a pseudo-second-order kinetic model. The isotherm results also showed that the modified lignin-based adsorbent exhibited monolayer adsorption. The adsorbent properties were mainly attributed to the incorporated amine functionalities as well as the increased specific surface area of the aminated CELF lignin.

INTRODUCTION

The demand for clean water is likely to increase driven by the rapid urbanization, expanding industrial activities, energy generation, and water pollution. Because of the limited fresh water resources on earth, this demand should be addressed by developing promising water purification techniques. The presence of organic dyes in industrial wastewater could cause some serious environmental concerns because of their poor biodegradability and toxicity to the exposed plants, living organisms, and even human being. As a result, these toxic dyes should be removed from the wastewater as much as possible before being discharged to land or water sources in an environmentally friendly manner. The textile industry consumes organic dyes, which represent 60% of the world’s dye consumption, and it was reported that 10−25% of water-soluble dyes is lost during the dyeing process, and 2−20% of dyes is released as the effluent into the water system after the dyeing process. Azo dyes are known as dyes containing $\equiv N\equiv N\equiv$ groups, representing 60−70% of commercially available dyes in the world. They are extensively used in the textile industry and become part of the textile effluents. Therefore, how to cost-effectively remove these toxic azo dyes from the industrial wastewater has become a topic of great interest. Adsorption is considered as an alternative method to the traditional combination of chemical and biological processes for the removal of dyes from aqueous solutions. Dye adsorption by various adsorbents is considered to occur primarily via $\pi$ interaction, hydrogen bonding, and electrostatic interactions. Several organic and inorganic materials including zeolite, lignocellulosic substrate, activated carbon, graphite, and graphene oxide have been all tested and shown to have different adsorption activities toward organic dyes in wastewater. Some of these adsorbents have a rather high dye-removal efficiency; however, low-cost renewable green bioadsorbents are still rare for this field of application, and further studies are urgently required. The biorefinery concept has received considerable attention in the last decade because of advances in biotechnology and
genetic engineering, offering a renewable and sustainable alternative to the production of common petrochemicals.14 Abundant lignocellulosic biomass is a second-generation nonfood feedstock that, when used by future biofinereries, has the potential to significantly offset the carbon footprint of the traditional refining.15 As one of the most important renewable fractions found in biomass, lignin is still significantly underutilized in the current biofinerery industries, which has mainly focused on transforming biomass carbohydrates to liquid fuels.16 It is anticipated that with the growing demand of biomass for production of fuels, the production of lignin would also substantially increase, potentially serving as a versatile platform for the production of biopolymers and renewable high-performance materials. The Renewable Fuel Standard (RFS) established in 2005 and further expanded in 2007 by the Energy Independence and Security Act (EISA) aims to ascend to 36 billion gallons of renewable fuel in 2022. Assuming a yield of 335 L per dry Mg of biomass, 223 million Mg of biomass will be used annually, producing about 62 million Mg of lignin.17 To avoid using lignin as a low-grade boiler fuel, new thermal and chemical processes are needed to generate value-added products from lignin.

A lignin macromolecule contains various amounts of functional groups including carbonyl, methoxyl, carboxyl, and hydroxyl groups, which offers promising opportunities to take advantage of its versatile functionality for multiple applications.18 Without any further chemical treatments, lignin could be directly incorporated into a polymeric matrix to be served as an antioxidant,19 a flame retardant,20 dye absorbent,21,22 and a UV stabilizer.23 Given the diversity of lignin, variability in functional groups (Figure 1).35 This provides a unique opportunity to assess the extent of lignin amination. Table 1 shows the effect of each experiment parameter on the extent of lignin amination by tracking the content of free H and G phenolic hydroxyl for each combination of factors (A: temperature, B: time, C: amine content, D: formaldehyde content, and E: acetic acid content) at four different levels was first applied to determine the optimal Mannich reaction conditions. The Mannich reaction can only occur between a high electron density carbon and an immonium ion formed from formaldehyde and an amine, thus the amine groups are expected to be introduced only at the ortho or para position of a phenolic hydroxyl group, converting H or G types of free phenolic hydroxyl group (or both) to C₃ substituted hydroxyl groups (Figure 1).35 This provides a unique opportunity to determine the extent of lignin amination by tracking the content of free H and G phenolic hydroxyl groups. Quantitative ³¹P NMR technique was used to track the content of free H and G phenolic hydroxyl groups, aiming to assess the extent of lignin amination. Table 1 shows the conversion of H and G types of free phenolic hydroxyl for each experiment of the designed orthogonal array. As indicated by Table 1, the effect of each experiment parameter on the extent of lignin amination increases in this order: acetic acid content (E) < reaction time (B) < DETA content (C) < formaldehyde content, and E: acetic acid content) at four different levels was first applied to determine the optimal Mannich reaction conditions. The Mannich reaction can only occur between a high electron density carbon and an immonium ion formed from formaldehyde and an amine, thus the amine groups are expected to be introduced only at the ortho or para position of a phenolic hydroxyl group, converting H or G types of free phenolic hydroxyl group (or both) to C₃ substituted hydroxyl groups (Figure 1).35 This provides a unique opportunity to

## RESULTS AND DISCUSSION

### Synthesis and Characterization of Adsorbents.

#### Orthogonal Experiments

A cosolvent enhanced lignocellulosic fractionation (CELF) method as a highly effective lignin-first pretreatment approach that is capable of extracting highly pure technical-grade lignin from corn stover. CELF applies aqueous mixture of tetrahydrofuran (THF) and dilute acid to greatly enhance the fractionation of lignin, hemicellulose, and cellulose fractions in biomass while promoting lignin fragmentation by limiting certain lignin condensation reactions typically suffered at high reaction severities. The obtained CELF lignin is depolymerized, containing lower aryl ether linkages and higher phenolic hydroxyl groups than the typical native milled wood lignin, cellulolytic enzyme lignin, or kraft lignin, which favors the subsequent amination process.31,32 The isolated CELF lignin was then aminated by diethylenetriamine (DETA) in the presence of formaldehyde under acid conditions via the Mannich reaction. An orthogonal array was also applied to test the optimal conditions for the Mannich reaction. The obtained final aminated lignin was thoroughly characterized by various analytical techniques including Fourier-transform infrared spectroscopy (FTIR), nuclear magnetic resonance spectroscopy (NMR), scanning electron microscopy (SEM), thermal gravimetric analysis (TGA), and Brunauer–Emmett–Teller (BET) surface area analysis. Finally, the performance of the obtained aminated CELF lignin as a biorenewable adsorbent for the removal of methylene blue (MB) and direct blue (DB) dyes was evaluated and compared to other reported adsorbents from literature. The combinatorial process takes advantage of the selectivity of the Mannich chemistry and the unique versatile functionality of CELF lignin such as low ether linkages and high phenolic OH content. It is fully expected that this study will provide a baseline for future studies to synthesize a renewable lignin-based dye adsorbent in advanced wastewater treatment systems.

### Figure 1

Mannich reaction between the phenolic G/S/H lignin and DETA, leading to the formation of phenolic C₃ substituted lignin units.

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content (D) < reaction temperature (A), according to the extremum of each factor (R value). The reaction temperature was found to be the most important factor, and the extent of amination appears to achieve its maximum at 75 °C. This could be because the Mannich reaction is an endothermic reaction thus it can be promoted by increasing the temperature. A further increase in temperature has been shown to have a negative effect on animation, which could be because of unnecessary formaldehyde and DETA evaporation, thus resulting in a decrease of reaction efficiency. In conclusion, the optimal combination parameters of the experiment are 75 °C, 4 h, 4 mmol DETA, 16 mmol formaldehyde, and 0.2 mL acetic acid according to the average values of each factor at different levels (K value). A large scale batch reaction was then performed at this optimal condition, and the obtained aminated CELF lignin was subsequently characterized by several state-of-the-art analytical techniques.

**FTIR Analysis.** The structural characteristics of the CELF lignin and aminated lignin were analyzed by FTIR as shown in Figure 2. Results showed that the aminated CELF lignin exhibited some basic adsorption peaks of CELF lignin, which indicates that the skeleton structure of lignin remains basically intact during the Mannich reaction. For example, hydroxyl group stretch (3400 cm⁻¹), asymmetrical stretching vibrations of −CH₂ and −CH₃ (2937 cm⁻¹), and symmetrical stretching vibrations of −CH₃ and −CH₂− (2844 cm⁻¹) are observed in both lignin samples. Nonetheless, there exist obvious differences between the lignin and aminated lignin samples. For example, the intensity of FTIR peaks associated with the aromatic C=H vibrations including 1603 and 1512 cm⁻¹ from the aminated lignin is significantly lower than that from the original CELF lignin. This is because the Mannich reaction mainly occurred in the aromatic region of lignin. In addition, the intensity of FTIR peaks associated with G units including 1267 cm⁻¹ (C=O stretch), 1113 cm⁻¹ (deformation vibrations of C–H), and 1030 cm⁻¹ (aromatic C–H in-plane deformation) and H units such as 1164 cm⁻¹ (C–O stretch) is decreased after amination reaction. On the other hand, the intensity of syringyl C–O stretch (1325 cm⁻¹) remains relatively strong after Mannich reaction. This is because the amine group could be only introduced at the C₆ or C₅ position of lignin and the C₅ position of the G units. Finally, a strong peak around 1650 cm⁻¹ representing the N–H bending vibrations in the amine structure (NH₂) appeared in the aminated lignin, validating the successful addition of the amine.

**2D HSQC Analysis.** Two-dimensional HSQC NMR has been comprehensively used in lignin characterization because of its versatility in offering structural insight into the lignin subunits and interlinkages. The reaction mechanism of Mannich reaction and the chemical structural transformation of CELF lignin during the amination reaction was further characterized by 2D HSQC NMR in this study. As shown in Figure 3, the CELF lignin possesses typical structural aromatic patterns of corn stover lignin. Peaks related to S, G, H, p-coumaric acid (pCA), ferulic acid (FA), and tricin (T) are all well defined in the aromatic region. Condensed S and G signals were also found in the CELF lignin, which are commonly observed from the lignin isolated by cosolvent pretreatment at temperature conditions of 180 °C or higher with acid. However, there exist dramatic differences between the original CELF lignin and modified lignin in both the aromatic and aliphatic regions. Specifically, it was found that the cross peaks associated with G and H lignin units were significantly altered during the Mannich reaction, while the signal of the S lignin units remained relatively stable. In addition, there are two Mannich reactive sites in tricin namely T₆ and T₈, which also disappeared in the aminated lignin because of the reaction of DETA and formaldehyde at these activate aromatic sites. Several intense signals were observed in

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**Table 1. L₁₆ (P₅)²⁴ Orthogonal Experiment Design of the Mannich Reaction between CELF Lignin and DETA under Acid Conditions**

<table>
<thead>
<tr>
<th>experiment</th>
<th>temp. (A)</th>
<th>time (B)</th>
<th>DETA (C)</th>
<th>formal. (D)</th>
<th>acid (E)</th>
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*For each run, 200 mg of lignin was dissolved in 2 mL of dioxane.

**Figure 2.** FTIR spectra of the corn stover CELF lignin and aminated CELF lignin.
the CELF lignin at $\delta_{C}/\delta_{H}$ 178/9.5–9.6 ppm and $\delta_{C}/\delta_{H}$ 123/7.5 ppm, representing the aldehyde (C$_{\alpha}$) and furanic C–H (C$_{3}$) signals of the five-substituted furfural derivatives, respectively. It has been reported that these types of furfural derivatives such as 5-hydroxymethylfurfural or 5-(methoxymethyl)furfural, which arose from sugar dehydration reactions could be condensed with the lignin structure during the acid catalyzed organosolv pretreatments. These types of structure were absent after the Mannich reaction possibly because of the reaction between aldehydes and primary amines to form Schiff bases.

In the aliphatic region, lignin interlinkages especially the $\beta$–O–4 linkages were dramatically cleaved during the CELF pretreatment process, and in fact, these linkages could be only detected at a noise level (data not shown). This is consistent with previous studies that reported CELF pretreatment performed at high severities (180 °C) was capable of achieving near-complete removal of its native $\beta$-aryl ether linkages without hydrogen input or further heterogeneous catalytic processing. This process is expected to generate a substantial amount of phenolic hydroxyl groups that favor the subsequent amination process. The methoxyl group

Figure 3. HSQC analysis of the original CELF lignin and its aminated product. (A) CELF lignin aromatic region; (B) aminated lignin aromatic region; (C) CELF lignin aliphatic region; (D) aminated lignin aliphatic region.
remains as the pronounced functional group in both lignin spectra. Compared to the unmodified CELF lignin, a significant amount of new signals appeared in the phenolic lignin side chain of aminated lignin, which was mainly ascribed to the methylene bridge of DETA introduced during the Mannich reaction. Our HSQC analysis also reveals that both the primary and secondary amines are capable of activating the formaldehyde. As shown in Figure 3, there still exist plenty of secondary amines in our proposed aminated lignin structure, which means that as new formaldehyde is activated by these protons, additional reactive phenolic G and H units could be subsequently grafted onto these partially aminated lignins until all the protons on the N atoms are replaced. A schematic diagram of Mannich reaction and the structural transformation of CELF lignin during the amination reaction is shown in Figure S3. The unmodified and aminated lignin was also subjected to a qualitatively visual ninhydrin test (Figure S4). The original CELF lignin had a negative/orange color indicating the absence of amines, while the change of color in the aminated lignin proved the existence of primary and secondary amine groups.

$^{31}$P NMR Analysis. Quantitative $^{31}$P NMR technique was further employed to determine different types of hydroxyl groups including aliphatic, phenolic, and carboxylic acid in the CELF lignin and aminated lignin, and the results are shown in Figure 4. The phosphorylation reaction of various OHs in lignin structural units with TMDP is shown in Figure S5. According to a recent study, the S hydroxyl group and condensed G hydroxyl groups are not fully baseline resolved and therefore are combined into C$_5$ substituted hydroxyl groups in this study to avoid any possible overestimation of S and underestimation of the G condensed unit. As compared to the original CELF lignin, a noticeable decrease in phenolic G and H hydroxyl groups was observed in the aminated CELF lignin after Mannich reaction. By contrast, the content of phenolic C$_5$ substituted hydroxyl groups including the S and condensed G and H units were considerably higher in the modified lignin than that in the original CELF lignin. The $^{31}$P results also indicated that the reactivity of the reaction sites in G lignin units was higher than that in the H lignin, and as a result, the proportion of the G unit decreased more obviously than the H unit. The slight loss of aliphatic hydroxyl group may result
from the possible loss of hydrophilic lignin fragment during the dialysis of aminated lignin. 

**SEM Analysis.** The modified lignin sample was obtained by rotatory evaporation and centrifugation followed by extended dialysis and freeze-drying. After repeated vacuum evaporation and centrifugation, the formation of a turbid suspension suggested the possible presence of aminated lignin nanoparticles. The morphological changes of CELF lignin during Mannich reaction are monitored by SEM and displayed in Figure 5. The unmodified CELF lignin in the solid state has a much larger particle size compared to the aminated lignin and appears granulated with irregular grains of compact structure. The surface of aminated lignin is much smoother than that of the original lignin. The nanospheric particles also aggregated into a micron-sized cluster with undefined shapes in the aminated lignin, which were possibly induced by the freeze-drying process. 

**Thermal Gravimetric Analysis.** The first derivative of the thermogravimetric (DTG) curves of the original CELF lignin and its aminated products exhibited different thermal degradation stages in N2 (Figure 6). Overall, the aminated lignin degraded faster than the unmodified lignin. In the pyrolysis range (200–600 °C), the DTG curve for the CELF lignin exhibits a single decomposition step with a decomposition peak temperature of 395 °C, while the curve for the aminated CELF lignin exhibits two composition steps, which are around 243 and 317 °C. The decomposition step above 300 °C is probably because of the cleavage of the C−C linkages and the demethoxylation of aromatic ring. The lower degradation temperature for aminated lignin is probably because of the lower C−N bond energy compared to the C−C bond. The decomposition step around 243 °C could be because of the degradation and evaporation of small molecular weight lignin fragments such as aliphatic side chains. The final degradation step around 600 °C is probably because of the crack of C−C/H bond of the charcoal that was formed during the pyrolysis. 

**BET Surface Area Analysis.** Table 2 summarizes the BET specific surface areas (SBET) and Barrett−Joyner−Halenda (BJH) pore volumes (VBJH) of the lignins before and after amination. Results indicated that the aminated lignin exhibited higher SBET and VBJH than those of the original CELF lignin. For example, the SBET values were 4.2 and 5.9 m²/g for the CELF lignin and aminated lignin, respectively. In addition, the mesoporosity of these lignin samples is also confirmed by the pore size distribution analysis (Figure S6). Because the aminated lignin had a larger surface area, it provided more adsorption sites, and hence enhanced dye removal can be anticipated.

**Removal of Dye by the Adsorbent.** To demonstrate the potential dye-adsorption property of the modified CELF lignin, two types of dyes were tested in the lignin-dye adsorption experiment: a cationic dye MB and an anionic azo dye DB 1. The qualitative and quantitative effect of lignin loading on the dye decolorization efficiency for the original and aminated lignin is shown in Figure 7. These results indicated that the aminated lignin showed drastically improved decolorization efficiency for both dyes especially the anionic DB 1 dye. For example, the amination process increased the DB 1 dye-removal efficiency from <5 to >90% even at extremely low lignin loadings. The decolorization efficiency for MB dye is proportional to the dose of lignin, while no correlation could be obtained between the efficiency of DB 1 dye removal and the concentration of lignin. This might be attributed to the unmodified lignin inability to adsorb DB 1 dye even at extremely high lignin dose and the aminated lignin strong ability to adsorb DB 1 dye even at low lignin loadings. The effect of amino content on the adsorptive property of the aminated CELF lignin was further investigated, and the results indicated that the dye decolorization efficiency was positively correlated to the amino content of the modified CELF lignin (Figure S7). The modified lignin has much higher decolorization efficiency toward the anionic dye (DB 1) compared to the cationic dye (MB). This could be mainly because of the electrostatic coupling between the cationic side chain (amine group) of the aminated lignin and the anionic sites of the DB 1 dye. It is well known that pH affects the adsorption of most organic pollutants as well as the surface charges of adsorbents. To further confirm that electrostatic interactions are key mechanism of adsorptive removal of dyes in DB 1 dye in aqueous solutions, the effect of initial pH on the dye decolorization efficiency and zeta potential of aminated CELF lignin was evaluated within the pH range of ~4.0 and 10.0 (Figure 8). At extreme acidic or basic conditions (pH < 3 or > 12), lignin samples were found to be partially or completely dissolved in aqueous solutions, thus their adsorption and surface charge behaviors were not investigated at these conditions. These results also indicated that high pH values resulted in a decrease in the adsorptivity of aminated CELF lignin, which could be because of the increase of the magnitude of the negative zeta potential as pH increases. The aminated CELF lignin has a point of zero charge (pHpzc) around 4.5, and its magnitude of zeta potential at each tested pH value (~2 to ~25 mV) is significantly lower than that of the unmodified CELF lignin ranging from ~68 to ~75 mV (Figure 8). This is because of the addition of the cationic amine group onto the side chain of aminated lignin. DB 1 dye has four sulfonate groups, thus it remains negatively charged at
basic conditions and even at highly acidic solutions as these protonated sulfonate groups have a $pK_a$ value lower than zero.\textsuperscript{53} Therefore, the increase of the net negative zeta potential could further cause a decrease of the electrostatic interactions between the cationic lignin side chain and the negatively charged DB 1 dye in aqueous solution. This suggests that electrostatic force is a major interaction for DB 1 dye adsorption at lower pH. Furthermore, the decolorization efficiency is still above 60% even at high pH, although there exists substantial electrostatic repulsion between the modified lignin and DB 1 dye. This indicates that mechanisms other than electrostatic interaction such as hydrogen bonding and $\pi-\pi$ stacking are also operative for the DB 1 dye adsorption on the aminated CELF lignin.\textsuperscript{54} On the other hand, hydrogen bonding, $\pi$-interaction, and limited electrostatic interaction between lignin dissociated carboxyl/hydroxyl groups and the cationic sites of the dye molecule are believed to be responsible for the MB dye adsorption.\textsuperscript{1} Figure 9 shows a proposed scheme for the MB and DB 1 dye binding to the unmodified CELF lignin and aminated lignin surface.

The Langmuir and Freundlich model was used to study the adsorption isotherms of azo-dyes, and their equations (eqs 1 and 2) are shown below

$$ Q = \frac{Q_m C}{K_c + C} \quad (1) $$

$$ \log Q = \log K_c + \frac{1}{n} \log C \quad (2) $$
where $c_e$ (mg/mL) is the equilibrium concentration, $Q_e$ (mg/g) is the equilibrium adsorption capacity, $Q_m$ (mg/g) is the maximum adsorption capacity of the Langmuir isotherm model, $K_L$ (mL/mg) is a Langmuir adsorption coefficient, $K_F$ (mL/mg) is the Freundlich constant, and $1/n$ is an indicator that reflects the nonlinear degree of adsorption. Figure 10 shows the Langmuir (A) and Freundlich (C) adsorption isotherms curves, and the linear analysis (B and D) indicated that the observed dye adsorption data for aminated CELF lignin were better described by the Langmuir isotherm model as confirmed by the higher coefficient $R^2$. The Langmuir fitting results suggested that the adsorption process between the lignin and the azo dye could be characterized as a monolayer type of adsorption. The Langmuir isotherm analysis indicated that the maximum adsorption capacity of the aminated CELF lignin is 502.7 mg/g for DB 1 dye. A direct comparison of the maximum adsorption capacities of different anionic azo dyes on various previously reported adsorbents is presented in Table 3. Based on our literature survey, it was found that the adsorption capacity of aminated CELF lignin is higher than that of all other adsorbents except carbon nanospheres. It should be noted that these adsorbents include commercial activated carbon, anion exchange membrane, and multiwalled carbon nanotubes, which are all well-known for their high aspect ratio, natural porosity, and strong ability to adsorb pollutants from the aqueous system. This suggests that the aminated CELF lignin, as a low-cost renewable resource, is highly suitable for the removal of azo-dyes from the aqueous solutions.

The rate of the adsorption process is an important factor that determines if the sorbent could be used on large scales in industrial applications and it can be determined by kinetic studies. In the study of solid–liquid static adsorption kinetics, the relationship between the time and the amount of adsorption is typically fitted through dynamic models. Figure 10. Adsorption isotherms of DB 1 by aminated CELF lignin. (A) Langmuir fitted adsorption isotherm curve; (B) linear fit of the Langmuir model ($R^2 = 0.99$); (C) Freundlich fitted adsorption isotherm curve; (D) linear fit of the La Freundlich model ($R^2 = 0.81$).

Table 3. Adsorption Performance of Different Adsorbents toward Azo-Dyes as Characterized by the Maximum Adsorption Capacity (mg dye/g Substrate)

<table>
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<th>adsorbent</th>
<th>dye adsorbate</th>
<th>maximum capacity (mg/g)</th>
<th>references</th>
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<td>9.1</td>
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</tr>
<tr>
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<td>90.8</td>
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<tr>
<td>lignin amine-coated Fe$_3$O$_4$</td>
<td>acid scarlet</td>
<td>176.5</td>
<td>54</td>
</tr>
<tr>
<td>chitosan</td>
<td>tartrazine</td>
<td>350</td>
<td>60</td>
</tr>
<tr>
<td>multiwalled carbon nanotube</td>
<td>DB 53</td>
<td>409.4</td>
<td>53</td>
</tr>
<tr>
<td>graphene oxide sponge</td>
<td>direct red 80</td>
<td>501.3</td>
<td>5</td>
</tr>
<tr>
<td>aminated CELF lignin</td>
<td>DB 1</td>
<td>502.7</td>
<td>present</td>
</tr>
<tr>
<td>chitosan-based hydrogel</td>
<td>erichrome black T</td>
<td>520</td>
<td>61</td>
</tr>
<tr>
<td>carbon nanospheres</td>
<td>acid red 88</td>
<td>555.6</td>
<td>62</td>
</tr>
<tr>
<td>Fe(OH)$_3$@cellulose hybrid fibers</td>
<td>Congo red</td>
<td>6897.3</td>
<td>63</td>
</tr>
</tbody>
</table>

mathematical models are usually adapted to analyze the dynamic models of adsorption process, namely pseudo-first-order (eq 3) and pseudo-second-order (eq 4) equation

$$\log(Q_e - Q_t) = \log Q_e - \frac{K_L t}{2.303}$$  (3)
the adsorption behaviors of azo-dye onto the aminated CELF lignin. This suggests that the pseudo-second-order kinetic model may be more suitable for describing the kinetics of the adsorption process. The pseudo-first-order and second-order equations, and their linear fitted plots and the parameters of the kinetic model are shown in Figure 11 and Table 4, respectively. It was found that the correlation coefficient ($R^2$) of pseudo-second-order equation kinetic model was higher than that of the pseudo-first-order model, suggesting that the pseudo-second-order kinetic model may be more suitable for describing the kinetics of the adsorption process of the DB 1 dye on the aminated CELF lignin. This suggested that the adsorption behaviors of azo-dye onto the modified lignin are dominated by chemical adsorption instead of diffusion process. It has been reported that the adsorption process is generally divided into three main stages, including the film diffusion stage, intraparticle diffusion stage, and the final actual adsorption stage. To further test if the intraparticle diffusion is the only rate determining step of the adsorption process, the intraparticle diffusion kinetic model is also fitted with the experimental data (Figure S9), and results indicated that the adsorption process has more than one controlling step. In addition, the relatively large boundary layer thickness as reflected by the large intercept of the linear plot ($t^{1/2}$ vs $q_t$) suggests that membrane diffusion might also have a great effect on the adsorption process.

Reusability of the absorbent represents an important aspect to minimize the cost of the overall adsorption process. The recycle performances of aminated CELF lignin for DB 1 dye removal were also investigated in this study. The results, as shown in Figure 12, show that no significant reduction in the adsorption efficiency is found for three cycles compared with that of the fresh adsorbent, although there is a gradual decrease in the dye-removal efficiencies possibly because of the incomplete of dye desorption. The dye-removal efficiency decreased to 65% for the fourth use, probably because of the saturation of the adsorbent surface. Thus, the recycle study demonstrated that the aminated CLEF lignin remained as an efficient adsorbent even after multiple reuses.

### Aminated CELF Lignin Characterization after Dye Adsorption

The FTIR spectra of aminated CELF lignin before and after dye adsorption are shown in Figure S10. After adsorption, two additional peaks at around 1200 and 1035 cm$^{-1}$ representing the stretching vibration bands of the sulfonate group appear in the aminated lignin because of the attachment of the DB 1 dye. This suggests that the lignin particles did not change dramatically and remained as an aggregated cluster with irregular shapes and heterogeneous surface, but their size appeared to increase significantly after dye adsorption. These observations clearly revealed that the DB 1 dye is adsorbed on the surface of the modified CELF lignin.

### Table 4. Parameters of the Adsorption Kinetic Model

<table>
<thead>
<tr>
<th>Kinetic Model</th>
<th>$q_e$ (mg g$^{-1}$)</th>
<th>$K_1$ (min$^{-1}$) or $K_2$ (g mg$^{-1}$ min$^{-1}$)</th>
<th>$R^2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pseudo-first order</td>
<td>285.8</td>
<td>0.01152</td>
<td>0.984</td>
</tr>
<tr>
<td>Pseudo-second order</td>
<td>511.7</td>
<td>0.00013</td>
<td>0.996</td>
</tr>
</tbody>
</table>

The observed experimental data were fitted to the pseudo-first-order and second-order equations, and the correlation coefficient ($R^2$) of pseudo-second-order equation kinetic model was higher than that of the pseudo-first-order model, suggesting that the pseudo-second-order kinetic model may be more suitable for describing the kinetics of the adsorption process of the DB 1 dye on the aminated CELF lignin. This suggests that the adsorption behaviors of azo-dye onto the modified lignin are dominated by chemical adsorption instead of diffusion process. It has been reported that the adsorption process is generally divided into three main stages, including the film diffusion stage, intraparticle diffusion stage, and the final actual adsorption stage. To further test if the intraparticle diffusion is the only rate determining step of the adsorption process, the intraparticle diffusion kinetic model is also fitted with the experimental data (Figure S9), and results indicated that the adsorption process has more than one controlling step. In addition, the relatively large boundary layer thickness as reflected by the large intercept of the linear plot ($t^{1/2}$ vs $q_t$) suggests that membrane diffusion might also have a great effect on the adsorption process.

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Development of low-cost renewable bioadsorbents for removal of toxic dyes from contaminated water has been a topic of great interest but remains challenging. In this study, the aminated corn stover lignin was synthesized via combinatorial CELF pretreatment and Mannich reaction to remove azo dyes from aqueous solutions. Under acid conditions, both the primary and secondary amines have high reactivity toward the H$_{135}$ and G$_5$ position of CELF lignin. SEM revealed that the original lignin particles are distributed in a large conglomerate, while the surface of aminated lignin becomes smoother, and the nanospherical particles of the modified lignin aggregate into nano- and micron-sized cluster with undefined shapes. The combination of CELF pretreatment and Mannich reaction significantly increased the adsorption behavior of aminated lignin toward azo DB 1 dye with a maximum capacity of 502.7 mg/g, which is significantly higher than that of many adsorbent materials reported in the literature. Recycle studied suggested that once recovered, the bioadsorbent was capable of maintaining a relatively high dye removal efficiency (>85%) even after three recycles. In conclusion, the proposed aminated CELF lignin could be considered as a cost-effective bioadsorptive platform for the efficient removal of azo dyes from aqueous solutions.

**Experimental Section**

**Feedstocks and Chemicals.** The Kramer corn stover was provided by the National Renewable Energy Laboratory (NREL, Golden, CO). The corn stover was knife-milled to pass through a 1 mm particle size interior sieve using a laboratory mill (model 4, Arthur H. Thomas Company, Philadelphia, PA). All the chemicals used in this study were used as received from Sigma-Aldrich without any further purification.

**Production of the CELF Lignin.** CELF pretreatment of corn stover was performed in a custom built 1 L Hastelloy Parr reactor (Parr instruments Company, Moline, IL) at 7.5 wt % solids loading and 0.5 wt % H$_2$SO$_4$ acid loading. The CELF reaction was sustained at 180 °C for 25 min in an equilvolum mixture of THF and water. After pretreatment, the reactor was quenched in a 25 °C water bath, and the liquid phase was separated from the pretreated solids through vacuum paper filtration. The CELF lignin was then isolated from the liquid phase by first neutralization with ammonium hydroxide followed by THF evaporation and subsequent vacuum filtration of the precipitated lignin from the neutralized liquor. The obtained CELF lignin was washed with water and diethyl ether and dried at 45 °C in an incubator. Once dried, the lignin was finally crushed to a fine powder in a mortar and pestle and stored in a container prior to further test and modification.

**Amination of the CELF Lignin.** The corn stover CELF lignin (≈200 mg) was mixed with 2 mL of dioxane in a round flask under constant stirring for 20 min at room temperature until the lignin was fully dissolved. Specified amounts of DETA, acetic acid, and formaldehyde were then added into the solution with continuous stirring. Formaldehyde was added stepwise to avoid unnecessary crosslinking reactions. Subsequently, the flask was heated in a sand bath, kept at a specified temperature (45, 60, 75, and 90 °C), and stirred for a specified time (1, 2, 3, and 4 h). Afterward, the reaction mixture was evaporated under reduced pressure to remove the majority of the organic solvent followed by dialysis with a molecular weight cut off of 1000 Da. The obtained aminated CELF lignin was finally freeze-dried and stored at room temperature before further characterization.

**Lignin Characterization before and after Amination.**

**FTIR Analysis.** The IR spectra were collected using a Spectrum One FTIR spectrophotometer (PerkinElmer, Wellesley, MA) equipped with a diamond-composite attenuated total reflectance cell from 1000 to 4000 cm$^{-1}$ with 128 scans at 4 cm$^{-1}$ resolutions.

**NMR Analysis.** NMR experiments were acquired with a Bruker Avance III HD 500 MHz spectrometer equipped with a 5 mm N$_2$ cryogenically cooled BBO H&F probe, according to previously published literatures. A standard Bruker pulse sequence (hsqcetgpspsi2.2) and an inverse-gated decoupling pulse sequence (Waltz-16) were applied for HSQC and $^3$P NMR experiment, respectively.

**SEM Analysis.** The morphology of lignin samples was observed with a scanning electron microscope (Zeiss Auriga, Germany) at an accelerating voltage of 5 kV. The samples were sputter-coated with Au using an SPI-Module sputter coater for 50 s. Imaging was subsequently captured at various magnifications from 2k to 20k.

**Thermal Gravimetric Analysis.** The TGA was performed by a TGA Q50 thermo-gravimetric analyzer (TA instruments, USA). Lignin samples (~5 mg) were loaded to a platinum sample pan (TA instruments) and heated in nitrogen from 25 to 105 °C at 20 °C/min. After incubating at 105 °C for 10 min, it was heated further from 105 to 800 °C at a heating rate of 20 °C/min.

**Zeta Potential Analysis.** The zeta potential of lignin suspensions was measured at different hydrogen ion concentrations while keeping a concentration of 1 mg/mL, using ZetaPALS (Brookhaven Instruments Corporation, NY). The mean zeta potential of each suspension was calculated from 10 measurements.

**Surface Area and Pore Size Analysis.** The N$_2$ adsorption—desorption measurement of samples was carried out at 77 K on a Quantachrome Autosorb iQ. The samples were first degassed at 353 K for ~17 h before being loaded into the analysis station. The pore volume and pore size distribution were determined using a BJH method. The specific surface area was calculated using BET in the P/P$_0$ range of 0.05~0.30.

**Dye Decolorization Study.** Various amounts of CELF lignin and aminated lignin were mixed with 25 mL of MB or

![Figure 13. SEM images of the aminated lignin after adsorption of DB 1 dye. (A) Mag. = 2k, (B) Mag. = 5k, (C) Mag. = 10k, (D) Mag. = 20k.](image)
DB 1 dye solution with a concentration of 50 mg/L. The mixture was left in an incubator at 25 °C and 150 rpm for 24 h. The concentration of dye in the supernatant of the solution at the equilibrium was determined by a UV spectrophotometer. The amount of dye adsorbed \((q_e)\) by lignin substrates was calculated based on the following eq 5

\[
q_e = \frac{v(c_0 - c_e)}{m}
\]

where \(c_0\) and \(c_e\) represent the initial and equilibrium concentrations of dye solution (mg/L), respectively, \(v\) is the volume of the total solution (mL), and \(m\) is the dry weight of the lignin sample (g). The maximum wavelength for MB and DB 1 dye was set at 663 and 624 nm, respectively. The extinction coefficient of MB and DB 1 dye was determined to be 149.3 and 12.3 L mol\(^{-1}\) cm\(^{-1}\) based on the Beer’s law calibration (Figures S1 and S2). The decolorization efficiency (\(\eta\)) is defined by

\[
\eta = \frac{c_0 - c}{c_0} \times 100\%
\]

where \(c_0\) represents the initial concentration, and \(c\) is the concentration of dye in the supernatant after decolorization. The adsorption isotherms of DB 1 dye onto aminated lignin were measured with varying concentrations of dye ranging from 0.05 to 1 mg/mL at 25 °C. To further investigate the kinetics of dye adsorption, the equilibrium concentrations of dye solution were measured from 5 min to 8 h after mixing 20 mg of solid with 25 mL of DB 1 dye solution with a concentration of 50 mg/L.

**ASSOCIATED CONTENT**

**Supporting Information**

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsomega.9b03717.

Calibration curve of dyes; schematic diagram of Mannich reaction; ninhydrin test of lignin amination; phosphitylation reactions between lignin OHs and TMDD; pore size distribution of lignin samples; effect of amino content on the adsorptivity of aminated CELF lignin; effect of contact time on adsorption; intraparticle diffusion kinetic of DB 1 dye; and FTIR spectra of the aminated CELF lignin before and after dye adsorption (PDF)

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**Author Contributions**

The experiments were performed through contributions of all the authors. All the authors have approved the final version of the manuscript.

**Notes**

The authors declare no competing financial interest.
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REFERENCES


